**SG marker co-localization with σNS**

* Seed cells 24 well plate containing 12mm glass coverslips 1 day before.
* Transfect cells with pCI S3 T3D using Lipofectamine 2000 according to manufacture’s instructions
* Add fresh media to cells 1 hour before the start of the experiment
* Change media and add Sodium arsenite (SA) (Sigma-Aldrich) at a final concentration of 1 mM dissolved in growth media for 1h
* Wash 1x with ice-cold PBS
* Fix with 3-4% PFA for 10 minutes at RT
* Incubate coverslips with staining buffer (SB, 0.05% saponin, 10 mM glycine, 5% FBS, and PBS) for 15 min at RT.
* Aspirate SB and incubate coverslips with primary antibody diluted 1:100 in 100 μl SB on rocker
  + Antibodies against σNS, TIA1, G3BP, Caprin1
* Aspirate primary antibody and wash coverslips X 1 with PBS.
* Incubate coverslips with secondary antibody diluted in SB for 1 h on rocker protected from light
* Aspirate secondary and wash coverslips X 3 with PBS, leaving in last wash
* Mount coverslips to glass slides with Prolong gold anti-fade reagent + dapi (6ul for a 12mm coverslip). Allow coverslips to cure overnight before imaging. Store slides in dark.